Intranasal delivery: An approach to bypass the blood brain barrier

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ABSTRACT

The blood brain barrier (BBB) represents one of the strictest barriers of in vivo therapeutic drug delivery. The barrier is defined by restricted exchange of hydrophilic compounds, small proteins and charged molecules between the plasma and central nervous system (CNS). For decades, the BBB has prevented the use of many therapeutic agents for treating Alzheimer’s disease, stroke, brain tumor, head injury, spinal cord injury, depression, anxiety and other CNS disorders. Different attempts were made to deliver the drug across the BBB such as modification of therapeutic agents, altering the barrier integrity, carrier-mediated transport, invasive techniques, etc. However, opening the barrier by such means allows entry of toxins and undesirable molecules to the CNS, resulting in potentially significant damage. An attempt to overcome the barrier in vivo has focused on bypassing the BBB by using a novel, practical, simple and non-invasive approach i.e. intranasal delivery. This method works because of the unique connection which the olfactory and trigeminal nerves (involved in sensing odors and chemicals) provide between the brain and external environments. The olfactory epithelium acting as a gateway for substances entering the CNS and peripheral circulation is well known. Also, it is common knowledge that viral infections such as common cold, smallpox, measles, and chicken pox take place through the nasopharynx. The neural connections between the nasal mucosa and the brain provide a unique pathway for the non-invasive delivery of therapeutic agents to the CNS. This pathway also allows drugs which do not cross the BBB to enter the CNS and it eliminates the need for systemic delivery and thereby reducing unwanted systemic side effects. Intranasal delivery does not require any modification of therapeutic agents and does not require that drugs be coupled with any carrier. A wide variety of therapeutic agents, including both small molecules and macromolecules can be rapidly delivered to the CNS using this method. The present review discusses the various applications, advantages and limitations of this novel approach.

KEY WORDS: CSF, nasal drug delivery, protein, polypeptide drugs

Introduction

Recent advances in the fields of pharmacology and molecular neurobiology have led to a greater understanding of disease processes, allowing development of new classes of therapeutic agents that can interact with specific intracellular and extracellular targets. Several drugs, peptides, biological response modifiers, and monoclonal antibodies are available and have proven value (a) in inhibiting a variety of malignant and infectious diseases; (b) in ameliorating neurotransmitter, enzyme, or growth imbalances in culture systems; (c) in animal models using direct intracranial administration. However, therapeutic efficacy in vivo, particularly with regard to the CNS is frequently diminished or nullified by the inability of the agent to reach and maintain effective concentrations in the brain for an appropriate length of time. Frequently, the molecule is too large or has polar functional groups and the blood brain barrier (BBB) limits its access to the CNS.

The BBB is a system of layers of cells at the cerebral capillary endothelium, the choroid plexus epithelium, and the arachnoid membranes, which are connected by tight junctions (zonulae occludens) and which together separate the brain and the cerebrospinal fluid (CSF) from the blood. These tight endothelial junctions can be 100 times tighter than junctions of other capillary endothelium. Thus, the barrier has many properties similar to a continuous cell membrane, allowing lipid-
soluble molecules transport across the membrane where hydrophilic solutes demonstrate minimal permeation.2 The BBB impedes the use, for example, of many of the newer genetically engineered drugs, such as humane recombinant neurotrophic factors and other therapeutic agents that can protect brain cells from damage and promote nerve repair. This impediment created by BBB can be overcome by three broad categories of techniques, namely

(i) Delivery across the BBB by manipulation of the drug to make BBB permeable to it (prodrug approach), utilization of carriers or transporters. However, these methods are complex and require drugs to possess certain specific characteristics and hence do not work effectively for all therapeutic agents. Drugs are administered by classical intravenous or intraperitoneal injections, or through the digestive tract, lung, or skin. However, part of these lipophilic chemicals are subject to being metabolized in the liver; resulting in a modification in the amount of circulating drug available to the brain. Moreover, the liver, kidney, intestine, skin, lung and also tissues separating the brain from the bloodstream, express enzymes able to metabolize xenobiotics. Another part of the dosage may be excreted by the kidney before entry into the CNS, rendering the precise amount of the drug that finally enters the brain difficult to be estimated. Similarly, drug delivery by intracerebroventricular (icv) devices (catheter, or osmotic pumps for intraventricular drug infusion) or by surgical implantation of devices that release an active molecule near its pharmacological target for variable time durations, has its own limitations.

(ii) Direct delivery to the brain by icv administration or surgical implantation, allows the administration of a precise amount of drug to the brain but requires invasive neurosurgery, hence is restricted to a limited number of applications (e.g., the administration of water soluble anticancer drugs, or treatment of intractable pain by direct central administration of morphine, to prevent fatal side-effects due to over-dosage).6 (Figure 1)

(iii) Bypassing the BBB: The third and the emerging approach is to bypass the BBB by intranasal delivery, which provides a practical, non-invasive, rapid and simple method to deliver the therapeutic agents to the CNS. This method works because of the unique connection between the nose and the brain that has evolved to sense odors and other chemical stimuli. This method is the thrust of this article.

Recently, Illum7 has thoroughly reviewed the possibilities, problems, and solutions of nasal drug delivery. She has reported that it is feasible to deliver challenging drugs efficiently such as small polar molecules, peptides and proteins and even the large proteins and polysaccharides like vaccines or DNA plasmids exploited for DNA vaccines. On the basis of clinical trials (Phase I and II) it is reported that the intranasal route is feasible for the transport of the drug to the CNS.

Intranasal delivery does not require any modification of the therapeutic agents and does not require that drugs be coupled with any carrier like in case of drug delivery across the BBB. A wide variety of therapeutic agents, including both small molecules and macromolecules can be successfully delivered, including to the CNS, using this intranasal method (Table 1).

Advantages

The advantages of intranasal delivery are considerable. This method is:

(1) Non-invasive, rapid and comfortable
(2) Bypasses the BBB and targets the CNS, reducing systemic exposure and thus systemic side effects
(3) Does not require any modification of the therapeutic agent being delivered
(4) Works for a wide range of drugs. It facilitates the treatment of many neurologic and psychiatric disorders
(5) Rich vasculature and highly permeable structure of the nasal mucosa greatly enhance drug absorption
(6) Problem of degradation of peptide drugs is minimized up to a certain extent
(7) Easy accessibility to blood capillaries
(8) Avoids destruction in the gastrointestinal tract, hepatic “first pass” elimination and gut wall metabolism, allowing increased, reliable bioavailability.

Limitations

(1) Concentration achievable in different regions of the brain and spinal cord, varies with each agent
(2) Delivery is expected to decrease with increasing molecular weight of drug
(3) Some therapeutic agents may be susceptible to partial degradation in the nasal mucosa or may cause irritation to the mucosa
(4) Nasal congestion due to cold or allergies may interfere with this method of delivery
(5) Frequent use of this route results in mucosal damage (e.g. infection, anosmia).

How does it work?

To understand the mechanism, pathways, distribution and absorption of therapeutic agents administered to the CNS by the intranasal route, a brief description of the nasal physiology is considered necessary.

Nasal physiology

The nose is divided into two nasal cavities via the septum.
The nasal cavity also has a key role in the sense of smell. The olfactory nerves, which originate as specialized olfactory nerve endings (chemoreceptors) in the mucous membrane of the roof of the nasal cavity above the superior nasal conchae, are the sensory nerves of smell. On each side of the septum nerve fibers pass through the cribriform plate of the ethmoid bone to reach the olfactory bulb where interconnections and synapses occur. From the bulb, a bunch of nerve fibers pass through the olfactory tract and reach the olfactory area in the temporal lobe of the cerebral cortex in each hemisphere, where the impulses are interpreted and odor is perceived. Another set of nerves emanating from the nasal cavity is the maxillary branch of the trigeminal nerves, which are general sensory nerves (Figure 2).

**Mechanisms**

Two mechanisms are involved in the nasal delivery, a fast rate that depends on lipophilicity, and a slower rate that depends on molecular weight. McMartin et al studied the transport of SS-6 (an octapeptide) and horseradish peroxidase through a rat’s nasal cavity. Their absorption studies are consistent with the non-specific diffusion of the penetrant molecules through aqueous channels located between the nasal mucosal cells, which impose a size restriction on nasal permeability. The data indicate that good bioavailabilities can be achieved for molecules up to 1000 Da (without enhancers) and good availability can be extended to at least 6000 Da with enhancers.

### Table 1

**Small molecules and macromolecules being studied for nasal delivery**

<table>
<thead>
<tr>
<th>Small molecules</th>
<th>Macro molecules</th>
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<tbody>
<tr>
<td>Adreno corticosteroids</td>
<td>Gentamicin, Cephalosporin, Penicillins, Tyrothricin</td>
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<tr>
<td>Antibiotics</td>
<td>Amino acids</td>
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<td>Antimigraine drugs</td>
<td>Peptides</td>
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<td>Antiviral drug</td>
<td>Calcitonin, Secretin, Thyrotropin-releasing hormone (TRH), Cerulein, Enkephalin analogs- Leucine enkephalin, Mekephamid Pentagastrin, SS-6, Substance P, Kyotorphin, Cholecystokinin</td>
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<tr>
<td>Cardiovascular drugs</td>
<td>Polypeptides and proteins</td>
</tr>
<tr>
<td>Central nervous system drugs</td>
<td>a. Albumins</td>
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<tr>
<td>a. Stimulants</td>
<td>b. Anterior pituitary hormones - Adrenocorticotropic hormone, Gonadotropin-releasing hormone, Growth hormone</td>
</tr>
<tr>
<td>b. Depressants</td>
<td>c. Biological products - Interferon, Vaccines</td>
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<tr>
<td>Autonomic nervous system drugs</td>
<td>d. Horseradish peroxidase</td>
</tr>
<tr>
<td>a. Sympathomimetics</td>
<td>e. Pancreatic hormones - Insulin, Glucagon</td>
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<tr>
<td>b. Parasympathomimetics</td>
<td>f. Posterior pituitary hormones - Oxytocin, Vasopressin</td>
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<td>c. Parasympatholytics</td>
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<td>Diagnostic drugs</td>
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<td>Histamine and antihistamines</td>
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<td>a. Histamine</td>
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<td>b. Antihistamines-</td>
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<td>c. Mast cell stabilizers -</td>
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<td>Narcotics and antagonists</td>
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<tr>
<td>a. Sympathomimetics</td>
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<td>b. Anticholinergics-</td>
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<td>c. Biological products -</td>
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<td>Sex Hormones</td>
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<td>a. Estrogens</td>
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<td>b. Androgens</td>
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<td>c. Adrenal corticosteroids</td>
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<td>d. Antimigraine drugs</td>
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<tr>
<td>e. Antihistamines-</td>
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<td>f. Posterior pituitary hormones -</td>
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<tr>
<td>Inorganic compounds</td>
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<td>a. Amino acids</td>
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<td>b. Nitrogen compounds-</td>
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<tr>
<td>c. Polypeptides and proteins</td>
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<td>d. Biological products -</td>
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<td>Vitamins</td>
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<td>a. Amino acids</td>
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<td>c. Polypeptides and proteins</td>
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<td>d. Biological products -</td>
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The transport mechanisms of different substances like insulin, mannitol or propranolol across the nasal mucosal tissue were studied by Wheatly et al. The transport of these substances occurs by a passive transport mechanism. The addition of deoxycholate (0.1%) reversibly increased the transepithelial conductance across the nasal membrane and enhanced the transport of mannitol and insulin. The transport of tyrosine and phenylalanine across rat mucosa was also studied by using an in-situ perfusion technique. It was found that both amino acids were absorbed by an active saturable transport process, which appeared to be Na⁺ dependent, and transport may have required metabolic energy as a driving force.

Water-soluble substances such as sodium cromoglycate are absorbed well and nasal absorption probably depends on aqueous channel diffusion (pores). The molecular size of the compound will be a determinant in the rate of absorption in such a channel.

**Drug distribution**

Drug distribution in the nasal cavity is an important factor that affects the efficiency of nasal absorption. The mode of drug administration may affect this distribution, which in turn can help determine the extent of absorption of a drug. Nasal deposition of particles is related to the individual's nasal resistance to airflow. With nasal breathing, nearly all the particles having an aerodynamic size of 10-20 mm are deposited on the nasal mucosa. The deposition of particles in the respiratory tract is a function of particle size and respiratory patterns. The density, shape, and hygroscopicity of particles, and the pathological conditions in the nasal passage will influence the deposition of the particle, whereas the particle-size distribution will determine the site of deposition and affect the subsequent biological response in animals and humans. Furthermore, improvement of the delivery system and drug formulation is necessary to achieve a better clinical effect and easier handling by patients.

Three mechanisms are usually considered in assessing particle deposition in the respiratory tract, i.e., inertia, sedimentation and diffusion, the first being the dominant mechanism in nasal deposition. Any particle with an aerodynamic diameter of 50 mm or greater does not enter the nasal passage. It was demonstrated that 60% of aerosolized particles of 2-20 mm are deposited in the anterior regions of the nostrils, 2-3 mm from the external nares. The site of drug deposition within the nasal cavity depends on the type of delivery system and the technique used in application.

**Drug absorption**

The first step in the absorption of drugs from the nasal cavity is passage through the mucus. Small, uncharged particles easily pass through this layer. However, larger or charged particles may find it more difficult to cross. Mucin, the principal protein in the mucus, has the potential to bind solutes, hindering diffusion. Additionally, structural changes in the mucus layer are possible as a result of environmental changes (i.e., pH and temperature). After a drug’s passage through the mucus, there are several mechanisms for absorption through the mucosa. These include transcellular or simple diffusion across the membrane, paracellular transport via movement between cells, and transcytosis by vesicle carriers. Obstacles to drug absorption are, potential metabolism before reaching the systemic circulation and limited residence time in the cavity.

Nasal absorption is affected by molecular weight, size, formulation pH, pKa of molecule, and delivery volume among other formulation characteristics. Molecular weight still presents the best correlation to absorption. The apparent cut-off point for molecular weight is approximately 1,000.
delivery in a variety of species. Neurotrophic factors such as AVP, CCK analog, and ADNF12 have been intranasally delivered to the brain in rodents. Hussain et al have demonstrated the therapeutic benefit of intranasal delivery of proteins in stroke studies. They have shown that intranasal IGF-I reduces infarct volume and improves neurologic function in rats with middle cerebral artery occlusion (MCAO). A preliminary report indicates that intranasal treatment is effective even when delayed for 4 h after the onset of MCAO. Gozes et al have shown that intranasal administration of a Vasoactive Intestinal Peptide Analog (VIP analog, containing 28 amino acids) prevented learning and memory impairments resulting from cholinergic blockade in rats treated with aziridinium. They also demonstrated that a nine amino-acid fragment of ADNF (ADNF-9) and an ANDF-like peptide (NAP) also protected against short-term memory loss in the same animal model.

Another technique aimed to increase nasal absorption is the utilization of bioadhesive agents. These compounds promote binding of drugs to biological material in the nasal cavity, thereby extending their residence time and allowing increased absorption. Common bioadhesive materials are carbopol, cellulose agents, starch, dextran, and chitosan.

Delivery of protein therapeutic agents /Macromolecules to CNS

In the age of advanced peptide, protein, and vaccine research, nasal administration of such compounds provides an attractive delivery route. In case of oral administration, the bioavailability of protein molecules tends to be relatively low due to their large molecular size and rapid enzymatic degradation. Because of their physicochemical instability and susceptibility to hepato-gastrointestinal “first pass” elimination, peptide/protein drugs are generally administered parenterally. It is on this background that intranasal administration seems a promising option. Most nasal formulations of peptide/protein drugs have been made up in simple aqueous or saline solutions with preservatives. Recently, more R&D work has been directed towards the development of nasal drug delivery systems for peptide/proteins. Currently, in the United States only four intranasal pharmaceutical products for systemic delivery have been marketed i.e. desmopressin (DDAVP), lypressin (Diapid), oxytocin (Syntocinon), and nafarelin acetate (Synarel).

Delivery of protein therapeutic agents to the CNS clearly involves extraneuronal transport as it occurs within minutes rather than hours. A number of protein therapeutic agents have been successfully delivered to the CNS using intranasal delivery in a variety of species. Neurotrophic factors such as NGF, IGF-I, FGF and ADNF12 have been intranasally delivered to the CNS in rodents. Studies in humans, with proteins such as AVP, CCK analog, MSH/ACTH and insulin have revealed that they are delivered directly to the brain from the nasal cavity. Hussain recently reviewed animal models to study nasal absorption and the effect of physicochemical and biopharmaceutical properties of drugs on the rate and extent of absorption. The review also discusses factors affecting peptide absorption and methods to improve the nasal bioavailability of peptides.

The bioavailability of protein molecules tends to be relatively low due primarily to their large molecular size and rapid enzyme degradation. As the number of amino acids increases beyond about 20, bioavailability becomes very low. To overcome these issues, much research has been conducted in the areas of absorption enhancers and bioadhesive agents. Absorption enhancers are used to increase the bioavailability. They are basically surfactants, glycosides, cyclodextrins and glycols. They improve absorption through many different mechanisms, such as increasing membrane fluidity, increasing nasal bloodflow, decreasing mucus viscosity, and enzyme inhibition. The classic example of a polypeptide compound with low nasal bioavailability is calcitonin. Its molecular weight is approximately 3,500 daltons and contains 32 amino acids in length. When calcitonin was given intranasally to rats and rabbits using a number of different cyclodextrins, its absorption as measured by a decrease in serum calcium concentration, was significant in comparison to the formulation without additive.

Research in humans has also provided evidence of direct delivery of macromolecules to the CNS from the nasal cavity. Kern et al have demonstrated CNS effects of intranasal corticotropin-releasing hormone (CRH) without altering plasma cortisol or CRH levels. Perras et al have reported that intranasal delivery of growth hormone-releasing hormone (GHRH) not only increased rapid eye movement sleep and slow wave sleep in humans, but also decreased growth hormone.

Al-Ghanameem et al carried out a study on the utility of the nasal route for delivery of 17β-estradiol, using its water-soluble prodrug. The study revealed that CSF concentration of 17β-estradiol following intranasal delivery of prodrug was higher compared to an equivalent intravenous dose. This result has a significant value in the treatment of Alzheimer’s disease.

The efficacy of peptide/protein delivered intranasally is highly dependent on the molecular structure of the drugs and their size. Respiratory epithelial cells are capable of absorbing peptide/protein by a vesicular transport mechanism, followed by transfer to the extracellular spaces, and subsequent uptake by the submucosal vascular network.

Delivery of DNA plasmids to the CNS

Of the several routes available for immunization, the nasal route is particularly attractive because of the ease of ad-
ministration and the induction of potent immune responses, particularly in the respiratory tract. However, adjuvants and delivery systems are required to enhance immune responses following nasal immunization. The use of microparticles [poly(lactide co-glycolide)] as adjuvants and delivery systems for protein and DNA vaccines for nasal immunization were reviewed by Vajdy et al.26 It has also been reported that after nasal administration of DNA plasmids, the level of plasmid in the brain was, 3.9 to 4.8 times higher than the plasmid concentration in the lungs and spleen. It was also found that the plasmid DNA reached the brain within 15 min following intranasal administration.77 The higher distribution of plasmid to the brain after intranasal administration indicates that nasal administration might be a potential route for the delivery of therapeutic genes to the brain with reduced side-effects in the other organs. The plasmid administered in this study was very large as was the plasmid detected in the brain.

Delivery of small molecules to the CNS

Many small molecules have been shown to be transported directly to the brain and/or CSF from the nasal cavity. This has been reviewed by Illum9 and Mathison et al.11 Anand kumar et al80 and David et al80 have demonstrated intranasal delivery of estrogen and progesterone respectively, to the CSF. Studies have also shown that drugs such as L-NAME89 and cocaine (at the lower end of the lipophilicity scale)91 have a higher CSF and olfactory bulb concentration after nasal administration than that obtained after parenteral administration. The properties of small molecules, including size and lipophilicity affect delivery to the CNS following intranasal delivery.82-85

Sakane et al90 reported that following intranasal administration of the antibiotic cephalaxin to rats, higher CSF concentration was reached at 15 min but it declined to approximately half that concentration at 30 min. Because cephalaxin does not cross the BBB well and because CSF concentration was 166-fold higher after intranasal administration than after systemic administration in spite of similar blood levels, it was concluded that cephalaxin entered the CSF directly from the nasal cavity. Using a series of fluorescein isothiocyanate-labeled dextrans (FITC-dextran) with increasing molecular weights, it was found that dextrans with molecular weights of up to 20,000 daltons could be transported directly from the nasal cavity of rats into the CSF.86 The concentration of the FITC-dextrans in the CSF increased with decreasing molecular weight. These FITC-dextrans are not found in the CSF after intravenous administration. Similarly, a comparison of the brain olfactory bulb concentrations achieved 30 min after intranasal administration of 7.4 n mol dopamine (153 daltons)86 with those obtained after intranasal administration of 7.4 n mol nerve growth factor (NGF) (26,500 daltons)86,17 to rats, revealed a five-fold higher delivery of the lower molecular weight dopamine. Comparing the percentages of the original dose remaining in the brain 30 to 45 min after intranasal administration of dopamine (0.12%)92 and NGF (0.023%)52 in rodents revealed a similar difference. In addition, with most small molecules, a significantly higher molar dose can be delivered intranasally than with larger protein or DNA therapeutic agents. Thus, considerably higher concentrations of small molecules are achievable in the CNS with intranasal delivery. Ishikawa et al93 reported that powder formulation of elcatonin utilizing CaCO3 improves the nasal bioavailability by increasing residence time in the nasal cavity and thus enhances the systemic bioavailability. Recently Bergstrom et al87 studied the uptake of picolnic acid (PA) in the brain. [3H]PA was administered via unilateral nasal instillation or i.v. injection to mice. Autoradiography demonstrated rapid uptake of radioactivity in the olfactory nerve layer and in the ipsilateral olfactory bulb following nasal instillation, which was maintained at a high level even after 4 h. On the other hand i.v. injection of [3H]PA demonstrated selective uptake and retention of radioactivity in the olfactory bulb. Hussain et al87 have found that intranasal administration of folic acid effectivly results in complete and very rapid absorption into the CNS. This provides a method of rapidly and reliably delivering folic acid, alone or in combination with other compounds, to the systemic circulation to produce a beneficial effect in the treatment or prevention of Alzheimer’s disease and stroke. Li et al80 reported rapid onset intranasal delivery of diazepam using ethyl-laurate-based microemulsion. At a 2 mg/kg dose, the maximum drug plasma concentration was arrived within 2-3 min, and the bio-availability (0-2 h) after nasal spray compared with i.v. injection was about 50%. The results suggest that this approach may be helpful during emergency treatment of status epilepticus.

Illum et al91 have studied the effect of chitosan-morphin nasal formulation vis-a-vis slow i.v. infusion of morphine in healthy volunteers who reported sedation at the earliest time point after nasal administration compared with i.v. administration. This suggests that after nasal administration morphine may be able to reach CNS more rapidly than after i.v. administration.

Conclusion

In summary, the advantages of intranasal delivery are considerable. It is both rapid and non-invasive. It bypasses the BBB and targets the CNS, reducing systemic exposure and thus systemic side effects. Even for drugs that can cross the BBB, it can reduce systemic side effects by reducing the need for the drug to enter the systemic circulation. It does not require any modification of the therapeutic agent being delivered and should work for a wide range of drugs. Intranasal delivery may facilitate the treatment and prevention of many different neurologic and psychiatric disorders.

References

Intrasenal delivery